

## *In vitro* micropropagation of *Eulophia andamanensis*–A tropical orchid

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**Abstract:** Rapid *in vitro* propagation of the tropical orchid *Eulophia andamanensis* was achieved by applying simple and reliable procedure for micropropagation. Pseudobulbs and axenic leaf buds of *Eulophia andamanensis* were placed on MS culture media with high auxin: cytokinin ratio. The incidence of moderate (~40%) contamination in cultures was reported due to source of explants. Further the shoot multiplication increased upon subculture on freshly prepared medium. The experiment cycles were repeatable. Media containing the growth regulators as 1 mg/l BAP + 0.5 mg/l NAA, 2 mg/l BAP + 0.5 mg/l NAA, 2 mg/l BAP + 1 mg/l NAA, 1 mg/l Kinetin + 1 mg/l NAA, 2 mg/l Kinetin + 1 mg/l NAA, 3 mg/l Kinetin + 0.5 mg/l NAA respectively. While the Pseudobulb explants on media MS + 1mg/l BAP + 0.5 mg/l NAA and 2mg/l BAP + 0.5 mg/l NAA and control only showed good response. Pseudobulb explants performed better than axenic buds. Callusing was not observed in leaf slices and buds explants.

**Keywords:** Pseudobulbs, monopodial orchids, flower stalks, micropropagation, vegetative regenerants

### INTRODUCTION

*Eulophia andamanensis* is one of the highly evolved terrestrial orchid species, known worldwide for its longest spike length (Recorded in Limca Book, 2013) and long lasting nature. It is called as green pretty orchid. The rare greenish white with maroon stripe flowers has made it exceptionally popular as a cut flower. Plants are commonly propagated from vegetatively by division of Pseudobulbs. To multiply through division, mother plants must be at least 3-4 years old and yield only very few preparative's Pseudobulbs and it grows very slow. To overcome these problems a reliable faster and successful multiplication for large scale commercial cultivation method for *Eulophia* is necessary. In orchids, shoot tip, meristem, leaf and leaf sections, stem segments, pseudobulbs, floral parts, aerial root etc, has successfully been used for *in vitro* propagation. Among these, pseudobulbs are used

as explants for large scale multiplication of commercial important *Eulophia andamanensis* orchids. A large number of growth hormones are used extensively for better regeneration and proliferation in orchids (Hasegawa *et al.*, 1985; Kim and Kako, 1982). Hence, the study was undertaken to find out the efficiency of growth hormones for *in vitro* proliferation, differentiation and growth of *Eulophia andamanensis* green pretty orchid.

Orchids are well known for its ornamental importance due its great diversity of flowers and long lasting in nature as well as therapeutic important for it's curing of several ailments by applying compounds of orchids (Vij *et al.*, 1984; 2000). The Orchidaceae family is one of the largest of flowering plants and they are terrestrial, epiphytic, or saprophytic herbs with about 1,000 genera and 25,000 species (Pridgeon, 1992; Hamilton, 1998 and Jain, 1980). Orchids are of

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enormous horticultural importance and also play a very important role for stability of the forest ecosystems (Kaushik, 1983). Orchid constitutes an order of monarchs in the world of ornamental plants (Stewart *et al.*, 1982). They are one of the most indulge plants and occupy top position among all flowering plants valued for cut flower production as potted plants. The propagation of orchids is naturally very difficult because of low viability of seeds and its different developmental stages to develop as adult plants (Morel 1964; Churchill *et al.*, 1973). For the conservation of endangered and medicinally important plant species, it is desirable to establish protocols for rapid clonal propagation (Kaur and Sarma, 1997; Bechtel *et al.*, 1980, Murashige *et al.*, 1962). The commercial propagation of the orchid very slow; to overcome to this problem in vitro propagation are needed for rapid production of healthy plants.

*Eulophia andamanensis* is an orchid endemic to the Andaman and Nicobar group of Islands are situated off the eastern coast of India between 6°45'2" N 93°12'2" E and 13°41'2" N 93°57'2" E. The genus *Eulophia* belongs to the subfamily Vandoideae, tribe Cymbidieae, and subtribe Cyrtopodiinae. *Eulophia andamanensis* is mostly found growing wild in the Little Andaman Islands, where the temperature ranges between 23–30° C and the annual average rainfall is 3473 mm. The growth habit is sympodial. The stem at the base is bulbous, with thick roots. The leaves are short during flowering, linear lanceolate. The bracts are shorter than the pedicel, the sepals 2 cm long, the lip shorter than the sepals. The sepals are linear lanceolate, 3–5 nerved, acuminate; both the sepals and petals are pale green in colour, the lip green at the base and white at the centre with maroon horizontal striations. The flowering period is from November to March, with the florets borne on long spikes (0.6–1.3 m long), and last for about 50 days.

In the present study, different culture media for direct shoot regeneration from nodes of pseudobulbs *Eulophia andamanensis* were tested in order to develop an efficient and rapid propagation method for these orchids.

## MATERIALS AND METHODS

The source plants of *Eulophia andamanensis* were maintained in Orchid house of Central Agricultural Research Institute, Port Blair. Juvenile shoots, pseudo bulbs and leaf dormant buds were used as the source of explants. The shoots were excised, defoliated and cut into 5-7 cm long pieces having three or four nodes. Explants were washed thoroughly in running tap water then washed with fungicide followed by surface sterilization under Laminar Air Flow hood with a 0.1% HgCl<sub>2</sub> solution for 10 minutes and rinsed thrice in sterile distilled water. The nodal explants of 1 cm length were cut aseptically and placed *in vitro* on MS medium containing 2% sucrose and supplemented with different combination and concentrations of plant growth regulators *viz.*, BAP, Kin and IAA. The pH of the medium was adjusted to 5.8 before adding 0.8% agar and autoclaved at 121°C for 15 minutes. The cultures were incubated at 25 ± 1°C under fluorescent light (2000 lux, 16 hr photoperiod).

## RESULTS AND DISCUSSION

The regeneration response in the explants was completely dependent to the use of growth regulators in the culture medium (Table 1). Culture initiation was affected by the physiological status of tissues besides the chemical stimulus in the nutrient pool. Explants, pseudobulbs and axenic leaf buds remained recalcitrant to regeneration responded to certain selected media combinations. The cytokinin, BAP (2.0 mg/l) proved to be beneficial for production and proliferation of multiple shoots (Fig1). The relative abundance of phenolics in *Eulophia andamanensis* and the negative effect of the phenolic oxidates on growth and differentiation of tissues in culture as evidenced from extensive browning of the medium and loss of cultures. (Ernst, 1974; 1985, 1994; Tanaka *et al.*, 1988). Excessive phenolic production is possibly due to increased polyphenol oxidase and catalase activity triggered by certain cultural conditions (Harvais, 1982). Phenolic exudations from the pseudo bulbs were observed in all the treatments. Browning of the medium, followed by leaf buds death and blackening of the leaf tips was common

**Table 1**  
Effect of different concentrations of plant growth regulators on in vitro development of *Eulophia andamanensis*.

| Sl. No. | Media                         | Explant       | No of explants inoculated | No of explants responded | % Response of explants |
|---------|-------------------------------|---------------|---------------------------|--------------------------|------------------------|
| 1.      | 1 mg/l BAP + 0.5 mg/l NAA     | Leaf cuttings | 25                        | Nil                      | 0                      |
|         |                               | Pseudo bulb   | 25                        | 6                        | 24                     |
|         |                               | Axenicbuds    | 25                        | 2                        | 8                      |
| 2.      | 2 mg/l BAP + 0.5 mg/l NAA     | Leaf cuttings | 25                        | Nil                      | 0                      |
|         |                               | Pseudo bulb   | 25                        | 7                        | 28                     |
|         |                               | Axenicbuds    | 25                        | 2                        | 8                      |
| 3.      | 2 mg/l BAP + 1 mg/l NAA       | Leaf cuttings | 25                        | Nil                      | 0                      |
|         |                               | Pseudo bulb   | 25                        | 4                        | 16                     |
|         |                               | Axenicbuds    | 25                        | Nil                      | 0                      |
| 4.      | 1 mg/l Kinetin + 1 mg/l NAA   | Leaf cuttings | 25                        | Nil                      | 0                      |
|         |                               | Pseudo bulb   | 25                        | 2                        | 8                      |
|         |                               | Axenicbuds    | 25                        | 3                        | 12                     |
| 5.      | 2 mg/l Kinetin + 1 mg/l NAA   | Leaf cuttings | 25                        | Nil                      | 0                      |
|         |                               | Pseudo bulb   | 25                        | 3                        | 12                     |
|         |                               | Axenicbuds    | 25                        | 2                        | 8                      |
| 6.      | 3 mg/l Kinetin + 0.5 mg/l NAA | Leaf cuttings | 25                        | Nil                      | 0                      |
|         |                               | Pseudo bulb   | 25                        | 3                        | 12                     |
|         |                               | Axenicbuds    | 25                        | 3                        | 12                     |
| 7.      | Control MS                    | Leaf cuttings | 25                        | Nil                      | 0                      |
|         |                               | Pseudo bulb   | 25                        | 6                        | 24                     |
|         |                               | Axenicbuds    | 25                        | 4                        | 16                     |



Figure 1: A fully matured with flowered spikes of *Eulophia andamanensis*.



Figure 2: Different stages of in vitro propagation of *Eulophia andamanensis*.

in all the cultures, which affected the survival rate of the plantlets.

Extensive use of growth regulators in the media selectively promotes the growth and proliferation of the explants. Growth regulators depending on type and concentration both inhibit and promote germination in orchids (Arditti *et al.*, 1981). The beneficial effects of NAA were in line with the results of several orchids (Chung *et al.*, 1983; 1998; 2003). NAA stimulated the seedlings in *Arundina graminifolia* (Kaur and Sarma, 1997) whereas, in

*Paphiopedilum spicerianum* and *Dendrobium chrysanthemum* auxins inhibited the growth of shoots and roots (Nayak *et al.*, 1997b; Vij and Kaur, 1998; Talukdar, 2001; Temjenasangba *et al.*, 2005).

## CONCLUSION

The results showed that the average regeneration of plantlets with well developed leaves on culture media. Moreover, medium with NAA/BAP gave rise to plantlets. Generative regenerants only arose directly from dormant buds. The multiplication factor of

generative regenerants on medium more than 50% was found significantly the best among all the treatments used. We managed to obtain direct regeneration without undesirable callus formation on successful culture media and thus to shorten the time needed for formation of first regenerants and reducing the possibility of the occurrence of somaclonal variability.

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## References

- Arditti J, Michaud JD, Oliva AP (1981), Seed germination of North America orchids. I. Native California and related species of Calypso, Epipactis, Goodyera, Piperia and Platanthera. *Botanical Gazette*, 142:442-453.
- Bechtel H, Cribb P, Launert E (1980), *Manual of cultivated orchid species*. MIT Press, Cambridge, Mass.
- Chang CA, Lee HH, Chen CC, Lin MJ, Wang CP (2003), Phytosanitary certification program of oncidium seedlings and its future the development of ornamental industry in Taiwan. *Plant Pathol. Bullet.* 12(3): 141- 148.
- Chung JD, Chun CK (1983), Asymbiotic germination of *Cymbidium ensifolium* I. Effect of basal media and growth regulators on germination of seeds and shoot emergence from rhizomes. *J. Kor. Soc. Hort. Sci.* 24: 236-242.
- Chung JD, Leu JH, Lee S, Kim CK (1998), Effect of medium composition on multiple shooting growth of mericlone from rhizome of shoot tip culture of temperate *Cymbidium* species. In: *Biological Abstracts*, 105(4): p. 50.
- Churchill ME, Ball E, Arditti J (1973), Tissue culture of orchids. I. Methods for leaf tips. *New Phytologist*, 72: 161-166.
- Ernst R (1974), The use of activated charcoal in a symbiotic seedling culture of *Paphiopedilum*. *Am. Orchid Soc. Bull.* 43: 35-38.
- Ernst R (1985), Seed and clonal propagation of *Phalaenopsis*. In *Proc. of 5<sup>th</sup> Asian Orchid Congress*, Ed. Rao, A.N. Singapore, pp31- 41.
- Ernst R (1994), Effects of thidiazuron on in vitro propagation of *Phalaenopsis* and *Doritaenopsis* (Orchidaceae). *Plant Cell Tissue and Organ Culture*. 39: 273-275.
- Hamilton R (1988), *When does it flower?* 2nd ed. Robert M. Hamilton, 9211 Beckwith Road, Richmond, B.C., Canada V6X 1V7.
- Harvais G (1982), An improved culture medium for growing the orchid *Cypripedium reginae* axenically. *Canadian Journal of Botany*, 51: 327-332.
- Hasegawa A, Ohashi H, Goi M (1985), Effects of BA, rhizome length, mechanical treatment and liquid shaking culture on the shoot formation from rhizome in *Cymbidium faberi* Rolfe. *Acta Horticulturae* 166: 25-40.
- Jain SK (1980), *Orchid and mountain flora of India*. 67th Session Indian Sci. Conger. Assoc., Calcutta.
- Kaur S, Sarma CM (1997), Selection of best medium for *in vitro* propagation of *Dendrobium Lindleyi* Steud. *Advances in Plant Sciences*, 10: 1-5.
- Kaushik P (1983), *Ecological and Anatomical Marvels of the Himalayan Orchids*. Today and tomorrow's printers and Publishers, New Delhi, India.
- Kim KW, S Kako S (1982), Effect of plant growth regulators on organ formation in the *Cymbidium* shoot apex culture *in vitro*. *Journal of the Japanese Society of Horticultural Science* 51: 106-114.
- Morel G (1964), Tissue culture - a new means of clonal propagation of orchids. *American Orchid Soc Bull.* 33: 473-478.
- Murashige T, Skoog F (1962), A revised medium for rapid growth and bioassay with tobacco tissue cultures. *Plant Physiol.* 15: 473 -497.
- Nayak NR, Rath SP, Patnaik S (1997b), In vitro propagation of three epiphytic orchids. *Cymbidium aloifolium* (L) Sw., *Dendrobium aphyllum* (Roxb.) Fisch. and *Dendrobium moschatum* (Buch- Ham) Sw. through thidiazuron-induced high frequency shoot proliferation. *Sci Hort.* 71:243-250
- Pridgeon A (1992), *The illustrated encyclopedia of orchids*. Timber Press, Portland, OR.
- Sheelavantmath SS, Murthy HN, Pyati AN, Ashok Kumar HG, Ravishankar BV (2000), *In vitro* propagation of the endangered orchid, *Geodorum densiflorum* (Lam.) Schltr. through rhizome section culture. *Plant Cell, Tissue and Organ Culture*. 60: 151-154.
- Stewart JWH, Linder E, Schlepe, Hall, A (1982), *Wild Orchids of Southern Africa*. Macmillan South Africa, Ltd., Johannesburg.
- Talukdar, A (2001), Multiple shoot induction in *Dendrobium aphyllum* Roxb. *J. Orchid. Soc. Ind.* 15: 35-38.
- Tanaka M, Kumura M, Goi M (1988), Optimal conditions for shoot production from *Phalaenopsis* flower stalk cuttings *in vitro*. *Sci Hort.* 35: 117-126.
- Temjensangba, S and Chitta RD (2005), Regeneration and mass multiplication of *Arachnis labrosa* (Lindl. Ex Paxt.) Reichb: A rare and threatened orchid. *Curr Sci.* 88: 1966-1969.
- Vij SP, Abhilasha D, Dhiman A (1994), Regenerative competence of *Bletilla striata* pseudobulb segments: a study *in vitro*. *J Orchid Soc.* 11(12): 93-97.

Vij SP, Pathak P(1989), Micropropagation of *Dendrobium chrysanthemum* Wall. through pseudobulb segments. *Journal of the Orchid Society of India* 3: 25-28.

Vij SP, Kher A and Ashish G (2000), Orchid micropropagation. In: Chadha KL, Ravindran P N, Saahijram Leela. (Eds.),

Biotechnology in Horticultural and Plantation Crops, 598-641. Malhotra Printing House, New Delhi, India.

Vij SP, Sood A, Pathak P (1989), On the utility of rhizome segments in micropropagating *Eulophia hormusjii* Duth. *J. Orchid. Soc. Ind.* 3: 41-45.