

The Effect of Water Stress on Improve of Drought Resistance in Armenian Apricot

Reza Kamrani*, Gagik Santrosian** and Naser Bouzari***

ABSTRACT: In this experiment the effect of water stress preconditioning was studied in one-year-old Armenian apricot (Hambar). Plants were submitted to different treatments including :

T-0 : control treatment.

T-1 : drip irrigated daily.

T-2 and T-3 : irrigated daily at 50% and 25% of T-0, respectively.

T-4 and T-5 : irrigated to field capacity every 3 and 6 days, respectively.

After 30 days, irrigation was withheld for 10 days, maintaining the T-0 treatment irrigated daily. After this period, the plants were re-irrigated to run-off and treated as control treatment. The stomatal closure and epinasty observed in response to water stress represented adaptive mechanisms to drought, allowing the plants to regulate water loss more effectively and prevent leaf heating. A substantial reduction in the irrigation water supplied combined with a high frequency of application (T-3 treatment) promoted plant hardening; the plants enduring drought better, due to their greater osmotic adjustment (0.77 MPa), which prevented severe plant dehydration and leaf abscission. Such a preconditioning treatment may be valuable for young apricot plants in the nursery stage in order to improve their subsequent resistance to drought. A 50% reduction in daily irrigation (T-2 treatment) did not significantly affect either gas exchange rates or leaf turgor, which suggests that water should be applied frequently if deficit irrigation is to be implemented.

Keywords: Gas exchange; Osmotic adjustment; Plant hardening; *Prunus armeniaca*; Water relations; Water stress.

Abbreviations : g_l : leaf conductance . LIA : leaf insertion angle . Pn : net photosynthesis . Tc-Ta : canopy to air temperature difference . TDR : time domain reflectometry . θ_v : volumetric soil water content . Ψ_m : soil matric potential . Ψ_{md} : mid-day leaf water potential . Ψ_0 : leaf osmotic potential . Ψ_{0s} : leaf osmotic potential at full turgor . Ψ_p : leaf turgor potential . Ψ_{pd} : pre-dawn leaf water potential.

INTRODUCTION

Apricot (*Prunus armeniaca* L.) belongs to family Rosaceae and among the angiosperms, it is one of the largest families having about more than three thousand species. Most apricot trees are cultivated in Mediterranean countries, under where drought periods are increasingly common, a fact which makes irrigation water the most limiting factor for apricot productivity, since it affects the viability of the young plantations. Plants have developed physiological responses as well as ecological strategies to cope with water shortages, either by stress avoidance or stress tolerance. These responses allow them to survive and even to maintain some growth under adverse conditions. Plant response depends on the nature of the water shortage inducing physiological responses

to short-term changes [1], acclimation to a different levels of water availability [2] and adaptation to drought [3, 4]. A knowledge of drought resistance mechanisms makes it easier to plan deficit irrigation strategies designed to save water while minimising the negative impacts on yield or crop revenue [5]. Previous studies have indicated that apricot drought resistance is mainly based on avoidance mechanisms, together with some degree of osmotic adjustment, when plants are submitted to short-term water stress periods [6]. However, drought imposition rates can have a large effect on the results of studies on drought resistance [7]. For these reasons, the aim of this study was to determine the ability of young apricot plants to drought hardening by the application of different water stress preconditioning treatments, as well as to

* Islamic Azad University, Bam Branch, Iran

** Seed and Plant Improvement Research Institute of Karaj, Iran

improve our understanding of the physiological mechanisms involved in the response of apricot plants to water stress. Such information may be valuable in the nursery stage in order to improve the drought resistance of young apricot plantations.

MATERIAL AND METHODS

Plant Material and Experimental Conditions

Hambar is an Armenian apricot. This experiment was carried out in an experimental garden in National Agrarian University of Armenian on one-year-old apricot trees, growing under field conditions in 40 litre pots (45 cm diameter) containing a mixture of clay loam topsoil and peat, with 6% organic matter. Pots were buried in the soil in order to reduce in soil temperature. Plants were drip irrigated daily using one emitter of 4 l h⁻¹ per tree, maintaining the soil matric potential at about -25 kPa that monitored with tensiometers placed at 15 cm depth. Fertilization was applied including: 66 g N, 49 g K₂O, 72 g P₂O₅ and 1.5 g Fe (per plant and year) through the drip irrigation system every 2 weeks. No root emergence from pots into the surrounding soil was observed. At the end of July 2010, 30 trees of similar appearance were selected, and the following preconditioning treatments were imposed: T-0 (control treatment) and T-1, drip irrigated daily to field capacity; T-2, daily irrigated at 50% of the control treatment; T-3, daily irrigated at 25% of the control treatment; T-4, irrigated every 3 days to field capacity and T-5, irrigated every 6 days to field capacity. Plants were maintained for 30 days under these irrigation conditions (preconditioning period), after which irrigation was withheld for 10 days in all plants, except the control treatment (T-0), which continued to be irrigated daily. The pots were sealed with plastic film and covered with soil in order to decrease the rate at which water stress developed. Then, all plants were re-irrigated to run-off and treated in the same way as the control treatment. During the preconditioning period maximum air temperature was 35 °C, with a vapour pressure deficit (VPD) of the atmosphere at mid-day of 4 kPa. During the stress period maximum air temperature was 28 °C and VPD 1.7 kPa. Photosynthetically active radiation (PAR) at mid-day, measured at the canopy surface with a line quantum sensor fluctuated around 1650 μmol m⁻² s⁻¹ during the experimental period. Soil and plant water status, and leaf gas exchange were measured every 6 days during the preconditioning period and every 2 to 3 days during the stress period. Volumetric soil water content were determined in 3 pots per treatment using

time domain reflectometry (TDR) equipment [8] and tensiometers at 15 cm depth, were determined. A pair of TDR probes was installed at a depth of 300 mm, midway between the tree trunk and pot border. Leaf water potential was measured at pre-dawn for one mature leaf per plant and 4 plants per treatment, using a pressure chamber, following the recommendations of Turner [9]. Leaves were fully expanded and were selected at random from the middle of the shoots. After measuring Ψ nitrogen and osmotic potential was measured after thawing the samples and extracting the sap, using a vapour pressure osmometer. Pre-dawn leaf turgor potential (Ψ_p) was derived as the difference between leaf osmotic and water potentials. Leaf osmotic potential at full turgor (Ψ_{os}) was measured on leaves adjacent to those used to measure leaf water potential. 4 leaves per treatment were taken at pre-dawn and rehydrated to full saturation, following the same methodology as for Ψ between the Ψ_{os}. Osmotic adjustment was estimated as the difference of stressed and control plants. Leaf conductance (g_l) and net photosynthesis (P_n) were measured at mid-day for a similar number and type of leaves as for leaf water potential, using a field-portable, closed gas-exchange photosynthesis system supplied with IRGA. Leaf was enclosed within a fan stirred one-litre chamber. The mean return flow rates of air circulating within the closed system and the leaf to air vapour deficit for all measurements were 280 μmol s⁻¹, and 1.8-2.4 kPa, respectively. Analyser was calibrated daily with two standard. The angle between leaf petiole and stem was measured with a transparent protractor to determine epinasty, the change in petiole angle. 10 randomised leaves per plant and 3 plants per treatment were measured. Canopy temperature was measured using an infrared thermometer at mid-day. 4 measurements were taken in 4 plants per treatment. Simultaneously, air wet and dry bulb temperatures were monitored. Defoliation was estimated by counting the number of leaves per plant at the beginning and at the end of the experimental period in 4 plants per treatment. The design of the experiment was completely randomised with 4 replications. One plant per replicate was used. Data were subjected to analysis of variance (ANOVA) procedures. Appropriate standard errors of the means (S.E.) and L.S.D.s at P = 0.05 were calculated.

RESULTS AND DISCUSSION

Preconditioning Period

0.05 range test. During the preconditioning period (Table 1) a substantial depletion in soil water was

observed in treatments T-3 (irrigated at 25% of control) and T-5 (irrigated every 6 days), with values of θ_v around 10.6%, and beyond the range of the tensiometer readings (< -80 kPa) (Table 1). Values of soil volumetric water content were slightly higher in pots from the T-2 treatment (irrigated at 50% of control) than those of the T-4 treatment (irrigated every 3 days). However, both promoted moderate plant water deficits, as indicated by the pre-dawn leaf water potential (Ψ_{pd}) values, which were around -0.7 MPa in these treatments. In the T-3 and T-5 treatments a more severe plant water deficit (Ψ_{pd} around -1.1 MPa) was registered. The values of leaf water potential at mid-day (Ψ_{md}) followed a similar behaviour to that of Ψ_{pd} (Table 1). Only the more severe water deficit situations (T-3 and T-5 treatments) induced decreases in leaf turgor potential ($\Psi_{treatment}$) (Table 1). values with respect to the control No significant differences in leaf osmotic potential at full turgor (Ψ) were found between treatments during the preconditioning period, with values of around -1.9 MPa in all the treatments. Similarly to that observed for leaf water potential, all the water-stressed treatments induced epinasty (a significant decrease in the leaf insertion angle values), particularly in the plants from the T-5 treatment (Table 2). A very strong relationship between LIA and Ψ_{md} was found, confirming the view .epinastic movements in apricot plants are dependent on plant water status [6]. Changes in leaf orientation allow to a lower incidence of solar radiation and, as a consequence, a reduction in water loss and leaf heating [10]. Leaf conductance (g_l) was reduced by the water deficits applied, except in the T-2 treatment, which showed similar values to the control treatment. A 50% of reduction in g_l values occurred in plants from the T-3 and T-5 treatments (Table 2). However, net photosynthesis (F_n) only decreased significantly in the severe water deficit treatments (T-3 and T-5) (Table 2). Both parameters were linearly correlated ($r = 0.82^{***}$, data not shown), which suggested a limitation in the photosynthetic capacity under water stress conditions [11]. Similar relationships between P_n and g_l have been reported for other *Prunus* species [12, 13], indicating an efficient co-ordination of stomatal behaviour and photosynthetic activity [14]. Within the range of mid-day leaf water potential values measured during the preconditioning period, a parallel decrease in leaf conductance and leaf water potential was found. The linear relationship indicates that the regulation of water losses occurred early and regularly [15, 16]. The lack of a critical threshold Ψ_{md}

to induce stomatal closure is similar to the findings of studies on almond [17], peach [18] and citrus [19], but contrast with those on apple [20] and senescent almond leaves [21]. Canopy temperature in control plants was around 31 °C, whereas plants from the T-3 and T-5 showed the highest canopy temperatures (around 35 °C). Canopy minus air temperature ($T_c - T_a$) values showed that well irrigated plants (T-1 treatment) keep their leaves 3 °C lower than air temperature (Table 2), which indicates the cooling effect of adequate transpiration levels [22, 23]. Stressed plants from the T-5 treatment presented a positive value of canopy minus air temperature (Table 2). In this sense, Ehler [24] indicated that $T_c - T_a$ values increase progressively when soil moisture is a limiting factor, for that reason it can be used as an index of plant water status [25].

Stress/recovery Period

At the end of the withholding period, soil water content values were near permanent wilting point (around 9.5%, data not shown) in all the stressed treatments. Plants which had not been preconditioned (T-1 treatment) and those from the treatment irrigated every 6 days during the preconditioning period $\Psi_{(T-5)}$ reached the highest plant water deficits ($\Psi_{pd} \sim -4$ Mpa). Plants from the treatment irrigated daily at 25% of the control (T-3) presented the lowest plant water stress, with values of ~ -1.6 MPa, compared to the -0.5 MPa measured in the control treatment (T-0). Leaf turgor potential was close to zero in all the plants after 10 days of withholding irrigation. Plants from the T-3 treatment presented the lowest decrease, with values of $\Psi_p = 0.7$ MPa, compared with the 1.4 MPa of the T-0 treatment. These plants also showed a smaller decrease in the leaf insertion angle. Water withholding induced a significant reduction in leaf osmotic potential at full turgor (Ψ_{os}) in all the preconditioned plants, with values of -2.16 , -2.49 , -2.04 and -2.22 MPa for T-2, T-3, T-4 and T-5 treatments, respectively. No significant differences were found in Ψ_{os} values between the control (T-0) and the unpreconditioned (T-1 treatment) plants, with values of -1.72 and -1.84 MPa, respectively. A higher amount of osmotic adjustment occurred in plants from the T-3 treatment (0.77 MPa). Osmotic adjustment may be responsible for maintaining of turgor in these plants [26]. In this sense, the obtained results confirmed that in apricot plants it is necessary to reach severe plant water deficits ($\Psi_{pd} < -2$ MPa) to trigger this tolerance mechanism [6]. Gebre and Kuhns [27] indicated that cottonwood plants submitted to water stress

preconditioning using different irrigation intervals developed a limited osmotic adjustment of 0.2 MPa, although this mechanism was not observed after severe water stress (10 days of withholding irrigation). Canopy temperature increased significantly in all the studied treatments, with values 3-4 °C above the air temperature (data not shown). Leaf conductance and net photosynthesis values were severely reduced by withholding irrigation in all the studied treatments. However, a smaller reduction in *g_l* and *P_n* was observed in plants from the T-3 treatment, which showed a 55% reduction with respect to the control treatment (T-0) values, compared with the 75% reduction observed in the rest of the stressed treatments. *g_h* Three days after irrigation was restored, pre-dawn leaf water potential reached similar values to those of the control treatment.

Leaf conductance recovery was rapid in plants from the T-5 treatment (3 days), whereas in T-2, T-3 and T-4, total recovery occurred 5 days after re-irrigation of plants, and two days later in plants from the unpreconditioned treatment (T-1). Net photosynthesis recovered more rapidly than *g_l*, reaching values close to those of control plants in all the stressed treatments on day 5 of the recovery period. The faster recovery in leaf conductance values after re-irrigation in plants of the T-5 treatment can be explained by the greater defoliation suffered by these plants. Most of the remaining leaves in these plants were young and have higher leaf conductance levels than mature leaves [18, 28, 29]. The relative delay in stomatal opening following rewatering, compared with the rapid recovery shown by Ψ , may be considered as a safety mechanism, which allows plants to regain full turgor more effectively [30].

Table 1

Volumetric soil water content (θ) potential at pre-dawn ($\Psi(\Psi_{ppdv})$, soil matric potential (Ψ) and at mid-day (Ψ_{mdm}), leaf water and leaf turgor potential) in the different water stress treatments during the preconditioning period.

treatment	θ (%)	Ψ_m (kPa)	Ψ_{pd} (Mpa)	Ψ_{md} (Mpa)	Ψ_p (Mpa)
T-0 (control)	28.46 d	-9.33 a	-0.46 c	-1.81 d	1.90 c
T-2 (50% T-0)	18.28 c	-39.60 b	-0.68 b	-2.44	1.78 bc
T-3 (25% T-0)	10.7 a	*	-1.08 a	-2.89 ab	1.44 ab
T-4 (every 3 d)	14.47 b	-55.25 c	-0.69 b	-2.62 bc	1.56 abc
T-5 (every 6 d)	10.40 a	*	-1.14 a	-3.18 a	1.23 a

Data are the average of 5 measurements taken every 6 days. Values followed by a different letter indicate the existence of significant differences

according to LSD0.05 test. *Beyond the range of the tensiometer.

Table 2

Leaf insertion angle (LIA), leaf conductance (*g_l*), net photosynthesis (*P_n*), and canopy to air temperature difference (T_c-T_a), in the different water stress treatments during the preconditioning period .

treatment	LIA	<i>G_l</i> (mmol m ⁻² s ⁻⁴)	<i>P_n</i> (μ molCO ₂ m ⁻² s ⁻⁴)	T _c -T _a (°C)
T-0 (control)	82.43 d	135.63 c	8.52 b	-2.75 a
T-2 (50% T-0)	62.50 c	109.02 bc	7.26 ab	-1.13 b
T-3 (25% T-0)	45.95 b	67.21 a	4.54 a	-0.4 bc
T-4 (every 3 d)	41.44 b	94.61 ab	6.57 ab	-1.20 b
T-5 (every 6 d)	31.4 a	63.86 a	4.36 a	0.35 c

Data are the average of 5 measurements taken every 6 days. Values followed by a different letter indicate the existence of significant differences according to LSD 0.05 test.

CONCLUSIONS

Young apricot plants exposed to slight-moderate water stress conditions developed avoidance mechanisms based on stomatal closure, accompanied by leaf epinasty, which can be considered as a complementary mechanism for regulating transpiration, and both have been recognised as important adaptive mechanisms to drought. Under

more severe water stress conditions ($\Psi_{pd} < -1.75$ MPa) partial defoliation occurred and osmotic adjustment was triggered as a tolerance mechanism. Water stress induced by daily irrigation at 25% of the control (T-3 treatment) promoted plant hardening. When these plants were submitted to severe water stress conditions, they showed a lower reduction in leaf water potential and gas-exchange parameters, as

well as lower epinasty, mainly due to their greater osmotic adjustment, which prevented severe plant dehydration and leaf abscission. This preconditioning treatment may be valuable in the nursery stage, since it hardens the plants against drought and so improve the survivability of the young apricot plantations. Also, from a comparative study of the tested treatments, we can conclude that, when lack of irrigation is to be applied, it is advisable to use a high frequency with reduced amounts of water than longer irrigation intervals, since neither gas exchange nor leaf turgor was reduced by this.

REFERENCES

- T.C. Hsiao, E. Acevedo, E. Fereres, D.W. Henderson. (1976), Water stress, growth and osmotic adjustment. *Philos. Trans. R. Soc. Lond.* 273, 479.
- E.D. Schulze. (1986), Carbon dioxide and water vapour exchange in response to drought in the atmosphere and in the soil. *Ann. Rev. Plant Physiol. Plant Mol. Biol.* 37. 247-274.
- J. Levitt. (1980), Responses of Plants to Environmental Stresses. Vol. II. Water, radiation, salt and other stresses, Academic Press, New York.
- R.G. Alscher, J.R. (1990), Cumming, Stress responses in plants: adaptation and acclimation mechanisms. *Plant Biology*, vol. 12, Wiley-Liss, New York. pp. 1-15.
- R. Domingo, M.C. Ruiz-Sánchez, M.J. Sánchez-Blanco, A. Torrecillas. (1996), Water relations, growth and yield of Fino lemon trees under regulated deficit irrigation. *Irrig. Sci.* 16, 115-123.
- A. Torrecillas, R. Galego, A. Pérez-Pastor, M.C. Ruiz-Sánchez. (1999), Gas exchange and water relations of young apricot plants under drought conditions. *J. Agric. Sci.* 132, 445-452.
- M. Martin, J. A. Morgan, G. Zerbi, D. R. Lecain. (1997), Water stress imposition rate affects osmotic adjustment and cell wall properties in winter wheat. *Ital. J. Agron.* 1, 11-20.
- G.C. Topp, J.L. Davis. (1985), Time-domain reflectometry (TDR) and its application to irrigation scheduling. *Adv. Irrig.* 3, 107-127.
- N.C. Turner. (1988), Measurement of plant water status by the pressure chamber technique. *Irrig. Sci.* 9, 289-308.
- M.J. Sánchez-Blanco, J.J. Alarcón, J. Planes and A. Torrecillas. (1994), Differential flood stress resistance of two almond cultivars based on survival, growth and water relations as stress indicators. *J. Hortic. Sci.* 69, 947-953.
- S.C. Wong, I.R. Cowan, G.D. Farquhar. (1979), Stomatal conductance correlates with photosynthetic capacity. *Nature* 282, 424-426.
- T.M. Yoon, H. Richter. (1990), Seasonal changes in stomatal responses of sweet cherry and plum to water status in detached leaves. *Physiol. Plant.* 80, 520-526.
- R.D. Harrison, J.W. Daniell, J.M. Chesire. (1989), Net photosynthesis and stomatal conductance of peach seedlings and cuttings in response to changes in soil water potential. *J. Am. Soc. Hortic. Sci.* 114. 986-990.
- M.M. Chaves, P.C. Harley, J.D. Tenhunen, O.L. Lange. (1987), Gas exchange studies in two portuguese grapevine cultivars. *Physiol. Plant.* 70. 639-647.
- A. Benzioni, A., R.L. Dunstone. (1990), Effect of air and soil temperature on water balance of jojoba growing under controlled conditions. *Physiol. Plant.* 74 (1988) 107-112. [16] R. Savé, J. Adillón, Comparison between plant water relations of *in vitro* plants and rooted cuttings of kiwi fruit. *Acta Hortic.* 282. 193-197.
- J.R. Castel, E. Fereres. (1982), Responses of young almond stress to two drought periods in the field. *J. Hortic. Sci.* 57, 175-187.
- P.C. Andersen, B.V. Brodbeck. (1988), Water relations and net CO₂ assimilation of peach leaves of different ages. *J. Am. Soc. Hortic. Sci.* 113, 242-248.
- R. Savé, C. Biel, R. Domingo, M.C. Ruiz-Sánchez, A. Torrecillas. (1995), Some physiological and morphological characteristics of citrus plants for drought resistance. *Plant Sci.* 110, 167-172.
- A.N. Lakso. (1979) . Seasonal changes in stomatal response to leaf water potential in apple. *J. Am. Soc. Hortic. Sci.* 104, 58-60.
- A. Torrecillas, M.C. Ruiz-Sánchez, A. León, A.L. García. (1988), Stomatal response to leaf water potential in almond trees under drip irrigated and non irrigated conditions. *Plant Soil* 112, 151-153.
- B.S. Sandhu, M.L. Horton. (1978), Temperature response of oats to water stress in the field. *Agric. Meteorol.* 19, 329-336.
- C.R. Sumayao, E.T. Kanemasu, T.W. Brakke. (1980), Using leaf temperature to assess evapotranspiration and advection. *Agric. Meteorol.* 22, 153-166.
- W.L. Ehrler. (1973), Cotton leaf temperatures as related to soil water depletion and meteorological factors. *Agron. J.* 65, 404-409.
- R.D. Jackson, R.J. Reginato, P.J. Pinter, S.B. Idso. (1979), Wheat canopy temperature: A practical tool for evaluating water requirements. *Water Resour. Res.* 13, 651-656.
- Kramer, P.J. (1983), Water Relations of Plants. Academic Press, New York, 489 pp.
- G.M. Gebre, M.R. Kuhns. (1993), Effects of water stress preconditioning on gas exchange and water relations of *Populus deltoides* clones. *Can. J. Forest Res.* 23, 1291-1297.

- M.C. Ruiz-Sánchez, R. Domingo, R. Savé, C. Biel, A. Torrecillas. (1997), Effects of water deficit and rewatering on leaf water relations of Fino lemon plants. *Biol. Plant.* 39, 623-631.
- J. Solárová, J. Pospíšilová. (1983), Photosynthetic characteristics during ontogenesis of leaves. 8. Stomatal diffusive conductance and stomata reactivity. *Photosynthetica* 17, 101-151.
- T.A. Mansfield, W.J. Davies. (1981), Stomata and stomatal mechanism. Paleg, D. Aspinall, (Eds.), *The Physiology and Biochemistry of Drought Resistance in Plants*, Academic Press, New York, pp. 315-346. ms, in: L.G.

