

Nematicidal activity of aqueous extracts of local plants against *Tylenchulus semipenetrans* Cobb 1912 and their Effects on Plant Growth Parameters of *Citrus jambhiri*

V. Bamel* and A. D. Huchche**

ABSTRACT: The nematicidal efficiency of aqueous extracts of neem (*Azadirachta indica*), Aak (*Calotropis procera*) and datura (*Datura alba*) on juveniles of *Tylenchulus semipenetrans* was evaluated at 3 doses (S, S/2, and S/4) and 3 exposure times (12, 24, and 48 h) in vitro. Neem gave the maximum larval mortality followed by aak and datura. The standard extracts were more toxic compared to diluted ones. Similarly, mortality increased with time of exposure. In the pot experiment, significant reductions were recorded in nematode populations and rate of multiplication in treatments receiving extracts as compared to control. The present study revealed the effectiveness of all tested plant extracts to root-knot nematodes without any chemical inputs.

INTRODUCTION

Citrus is the India's second most important commercial fruit in terms of area and production after mango. Amongst different microorganisms like fungi, bacteria, viruses and nematodes which are the important agents affecting the quality and quantity of citrus, nematodes play a key role.

Plant parasitic nematodes present some of the most difficult pest problems encountered in our agricultural economy. Among nematodes, citrus nematode *Tylenchulus semipenetrans* is causing immense damage to citrus trees^[2]. This nematode occurs throughout the world citrus growing areas and causes a serious disease known as slow decline. Affected trees exhibit reduced vigor, chlorosis, leaf fall, die back and reduced production and quality of fruit^[4]. The nematode (*Tylenchulus semipenetrans*) is the primary cause of citrus decline. The nematode never kills trees but growth and yield of infected trees slowly decline. Severity of the disease can vary from minor to severe decline of infected trees. Symptom expression is influenced by several physical and chemical factors as shown by several investigations^[5].

The use of chemicals for nematode control on

large scale is an expensive and impracticable operation. This situation demands the search for cheaper alternative control measure which can be made available to small growers.

There are reports that certain plant parts and extracts possess nematicidal properties^[6-8]. Application of the plant parts or extracts to nematode infested soil affects nematode directly and stimulates soil microbes that reduce nematode populations^[9,10].

Under this context, the use of plant extracts with nematicidal properties can prove to be effective, cheaper and safer control measure.

MATERIALS AND METHODS

Leaves of datura (*Datura alba*), neem (*Azadirachta indica*) and Aak (*Calotropis procera*) were washed under running water and chopped. Twenty five grams of chopped leaves of each test plant were then macerated in an electric blender and soaked separately in 100 mL of distilled water. After 24 h, samples were filtered through Whatman filter paper No.1. The extracts were arbitrarily termed as standard "S" and subsequent dilutions viz., S/2 and S/4 were prepared by the addition of requisite quantities of distilled water.

* Division of Nematology, Indian Agricultural Research Institute, New Delhi, India - 110 012

** Central Citrus Research Institute, Amravati Road, Nagpur, Maharashtra, India - 440 033

* Corresponding author E-mail: vbamel@yahoo.com

In order to evaluate the effect of different water extracts on the larval mortality of *Tylenchulus semipenetrans*, 1 mL of nematode suspension containing about 30 juveniles was poured into the petri dish with the help of pipette and 5 mL of the extract was added in the petri dish with the help of pipette. The same procedure was repeated for all the extracts. Petri dishes containing distilled water served as control. Each treatment was quadruplicated. The petri dishes were placed at room temperature which ranged from 25 to 30°C. Dead and surviving nematodes were counted after 12, 24 and 48 h. Mortality was assessed by touching the nematodes with fine needle and percentage of larval mortality was calculated by the following formula:

$$\% \text{ Mortality} = \frac{\text{No. of larvae killed}}{\text{Total no. of larvae}} \times 100$$

The effect of the standard extracts of the leaves of the test plants was then studied in pots. For this purpose three months old seedlings of highly susceptible citrus cultivar Jatti khatti (*Citrus jambhiri*) were transplanted singly in pots (15x12 cm) containing formalin sterilized sandy loam soil. After one week of transplanting, the plants were inoculated with approximately 5000 juveniles/pot of *T. semipenetrans*. Standard extracts of leaves of each of datura, neem and Aak @ 100 mL/pot were applied after five days of inoculation. Each treatment was replicated five times. After three months, plants with soil were gently removed from pots and their roots were carefully washed under running tap water. Data were recorded on plant height, fresh and dry weights of shoot and root, final population of nematodes in soil, number of females per gram of root and rate of multiplication. All the data recorded were analyzed statistically by using analysis of variance and means were compared by using least significant difference test at 5% level of probability^[11].

Results

Analysis of variance regarding plant extracts, time of exposure, concentrations and their interaction was highly significant. Comparison of treatment means of leaf extracts of test plants indicated that neem gave the maximum mortality followed by datura and Aak as shown in Table 1. Similarly comparison of treatment means regarding time of exposure revealed that the maximum mortality was observed after 48 h of exposure followed by 24 and 12 h (Table 2). Further, it is evident from the comparison of

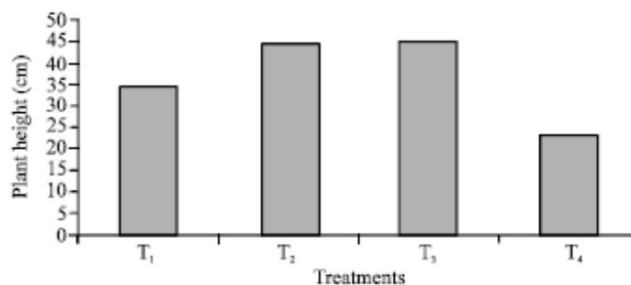
means (Table 3), that standard concentrations of the leaves extracts of test plants showed maximum mortality as compared to other concentrations. Mortality in distilled water was negligible. Mean individual percent mortalities in leaf extracts of test plants at different concentrations and exposure times are given in Table 4.

When standard leaf extracts of test plants were applied in pots, plant height and fresh and dry weights of shoots and roots were greater. Comparison of treatment means indicated that plant height was the maximum in Aak and neem extracts followed by datura extract showing 98.86, 96.15 and 49.91% increases over control (Fig. 1).

Comparison of treatment means revealed that fresh and dry shoot weights were maximum in neem extract which showed 200.14 and 223.63% increases over control, followed by Aak and datura extracts showing 187.85 and 196.63, 112.29 and 126.97% increases over control, respectively (Fig. 2 and 3).

Similarly, it is obvious from the comparison of treatment means that fresh and dry root weights were maximum in neem extract giving 205.71 and 122.52% increases over control and increases in these parameters in case of Aak and datura extracts were 166.03 and 65.76%, 62.85 and 18.91% over control, respectively (Figs. 4 and 5).

On the other hand significant reductions were recorded in nematode populations and rate of multiplication in treatments where extracts were applied as compared to control.



T₁ = Datura extract T₂ = Neem extract
T₃ = Aak extract T₄ = Control (Nematode alone)

Fig. 1. Effect of plant extracts on plant height (cm)

Table 1. Effect of plant extracts on larval mortality of *Tylenchulus semipenetrans*

Plant extracts	% mortality
Neem	19.44 ^a
Aak	14.78 ^b
Datura	17.23 ^b
LSD value	0.3796

Means were separated by LSD test at 5 % level

Table 2. Effect of different exposure times on larval mortality of *Tylenchulus semipenetrans*

Exposer time (h)	% mortality
48	19.02 ^a
24	17.09 ^b
12	15.25 ^c
LSD value	0.3796

Means were separated by LSD test at 5 % level

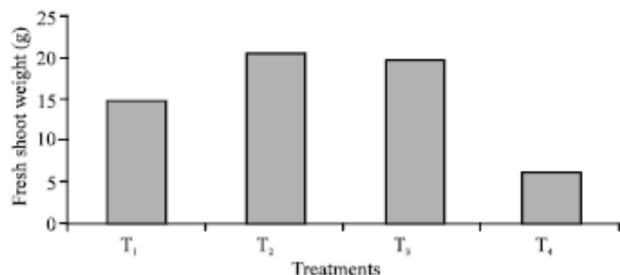
Table 3. Effect of different concentrations on larval mortality of *Tylenchulus semipenetrans*

Concentrations	% mortality
S	28.06 ^a
S/2	22.30 ^b
S/4	18.00 ^c
Distilled water	0.25 ^d
LSD value	0.4383

Means were separated by LSD test at 5 % level

Table 4. Effect of leaf extracts of test plants on larval mortality of *Tylenchulus semipenetrans* (Means of 4 replications)

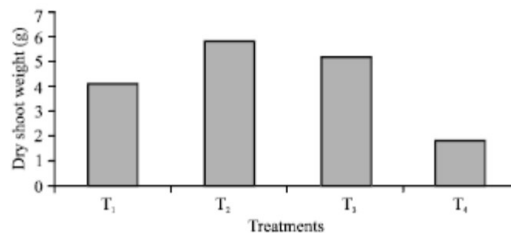
Plants	Concentrations	Percent mortality after		
		12 h	24 h	48 h
Datura	S	100	100.00	100.00
	S/2	75.55	81.10	96.66
	S/4	54.44	81.10	86.66
	DW	1.11	1.11	2.21
Neem	S	100.00	100.00	100.00
	S/2	10.00	100.00	100.00
	S/4	69.44	84.44	89.99
	DW	0.00	0.00	1.11
Aak	S	100.00	100.00	100.00
	S/2	57.77	71.10	87.8
	S/4	45.55	58.88	69.99
	DW	0.00	1.11	1.11



T₁ = Datura extract T₂ = Neem extract
T₃ = Ak extract T₄ = Control (Nematode alone)

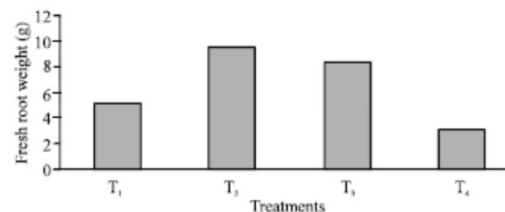
Fig. 2. Effect of plant extracts on fresh shoot weight (g)

Final population of nematodes in soil, number of females per gram of root and rate of multiplication was minimum in case of neem extract showing 78.72, 70.24 and 78.68% reductions over control, respectively. Aak extract was equally effective as neem extract giving 71.74, 60.57 and 71.82% decreases over control.



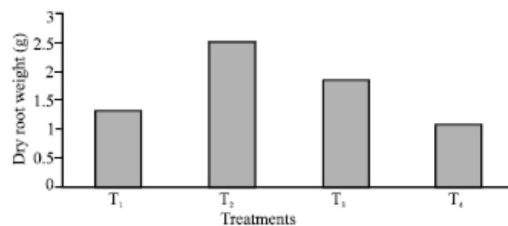
T₁ = Datura extract T₂ = Neem extract
T₃ = Ak extract T₄ = Control (Nematode alone)

Fig. 3. Effect of plant extracts on dry shoot weight



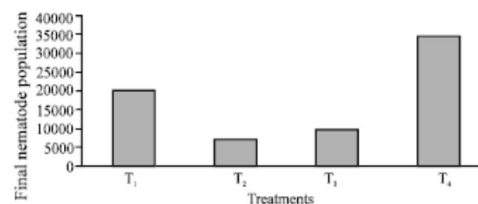
T₁ = Datura extract T₂ = Neem extract
T₃ = Ak extract T₄ = Control (Nematode alone)

Fig. 4. Effect of plant extracts on fresh root weight



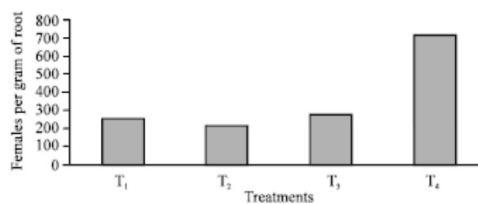
T₁ = Datura extract T₂ = Neem extract
T₃ = Ak extract T₄ = Control (Nematode alone)

Fig. 5. Effect of plant extracts on fresh dry weight



T₁ = Datura extract T₂ = Neem extract
T₃ = Ak extract T₄ = Control (Nematode alone)

Fig. 6. Effect of plant extracts on final nematode population



T₁ = Datura extract T₂ = Neem extract
T₃ = Ak extract T₄ = Control (Nematode alone)

Fig. 7. Effect of plant extracts on no. of females/ gram of root

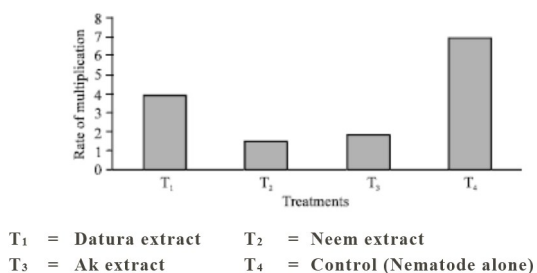


Fig. 8. Effect of plant extracts on rate of nematode multiplication

Datura extract was significantly different from neem and Aak extracts in case of final population of nematodes in soil and rate of multiplication while it was equally effective as neem and Aak extracts in case of number of females/g of root showing 44.50, 64.22 and 44.49% reductions in these parameters over control (Figs. 6-8)

DISCUSSION

When the water extracts of neem, Aak and datura and their subsequent dilutions were studied on the larval mortality of citrus nematode in laboratory, significant results were achieved. The nematicidal properties of these plants were tested in laboratory^[6,7,12,13]. The mortality of the larvae might be attributed to the chemicals contained in the extracts. These chemicals might have penetrated directly and inhibited acetyl cholinesterase and other esterases like cholinesterase enzyme. Hydrolysis of acetyl choline by acetyl cholinesterase is a vital part of neurotransmission in the nervous system. Cholinesterase and esterases also function in various metabolic systems. The existence of acetyl cholinesterase in plant parasitic nematodes was first demonstrated by using a site specific acetyl thiocholine substrate reaction that was inactivated by known cholinesterase inhibitors^[14].

It was suggested that the inhibition of body activity may result from acetyl cholinesterase inhibition that was irreversible at high concentrations due to continued binding of the enzymes^[15]. Various other effects observed include the prevention of or decreased mobility and delay in the molting processes. Many of these behavioral changes may relate to inhibition of various esterases enzymes.

When the standard extracts of leaves of the test plants were applied, plant height, fresh and dry weights of shoot and root were significantly more as compared to control. The reproduction factor was significantly low in pots where neem extract was applied. The findings are in line with those of other

workers^[7,16-19].

The protective action of the leaf extracts of these plants against citrus nematodes appears to be due to the presence of chemicals and growth stimulators in them. Possibly these chemicals are either absorbed by the root or an elicitor/activator reaction was initiated by some factor present in the extracts^[20,21]. According to Kast^[22] an induced defense mechanism may have some practical utility in integrated biological nematode management strategies. The root improvement could be attributed to poor penetration and later retardation in different activities of 2nd stage juveniles such as feeding and/or reproduction as suggested by Bunt^[23].

From the present as well as from the earlier studies^[24,25], it would appear that leaves of neem possess some broad spectrum factor which has the potential against nematodes with different modes of feeding. Systemic action of azadirachtin, a triterpenoid from neem, against citrus nematode has been reported^[26]. The effective nematicidal activity of neem leaf extract might have been due to the action of its active principles viz., Nimbine, Nimbinine, Nimbidine, Thionemone and Margosinone^[27].

These results indicate that extracts of these **medicinal plants** can prove helpful in the control of plant parasitic nematodes. Application and use of plant extracts will probably be easy and economical as compared to chemical treatment. The active principles of these medicinal and poisonous plants, if extracted in pure form in large quantities, would probably form the base for the development of indigenous nematicides.

REFERENCES

1. Milne, D.L., 1977. The impact of new nematicide and irrigation practices on method of citrus nematode control. Proc. Int. Soc. Citric., 3: 835-838.
2. Baines, R.C., O.F. Clark and W.P. Bitters, 1948. Susceptibility of some citrus and other plants to citrus root nematode *Tylenchulus semipenetrans*. Phytopathology, 38: 912-912.
3. Cohn, E., 1969. The citrus nematode *Tylenchulus semipenetrans* Cobb) as a pest of citrus in Israel. Proc. First Int. Citrus Symp., 2: 1013-1017.
4. Gundy, S.D.V., J.P. Martin and P.H. Taso, 1964. Some soil factors influencing reproduction of the citrus nematode and growth production of sweet orange seedlings. Phytopathology, 54: 294-299.
5. Nandal, S.N. and D.S. Bhatti, 1983. Preliminary screening of some weeds shrubs for their nematicidal

- activity against *Meloidogyne javanica*. Ind. J. Nematol., 13: 123-127.
6. Awan, M.N., N. Javed, R. Ahmad and M. Inam-ul-Haq, 1992. Effect of leaf extracts of four plant species on larval mortality of citrus nematode *Tylenchulus semipenetrans* Cobb) and citrus plant growth. Pak. J. Phytopathol., 4: 41-45.
 7. Sharma, R. and P.C. Trivedi, 1992. Effect of root extract of some plants on larval hatching of *Meloidogyne incognita*. Curr. Nematol., 3: 31-34.
 8. Nandal, S.N. and D.S. Bhatti, 1986. Influence of four plant extracts on the hatching of *Meloidogyne javanica* and invasion of host roots. Nematol. Medit., 14: 291-294.
 9. Reddy, P.P., M.S. Rao and M. Nagesh, 1996. Management of citrus nematode, *Tylenchulus semipenetrans* by integration of *Trichoderma harzianum* with oil cakes. Nematol. Medit., 24: 265-267.
 10. Steel, R.G.D. and J.H. Torrie, 1980. Principles and Procedures of Statistics. 2nd Edn., McGraw Hill Book Co., New York, pp: 173-177
 11. Lee, M.J., 1987. The effect of extract of *Melia azadirach* on *Meloidogyne incognita*. Q. J. Chine. For., 20: 1-7.
 12. Mani, A., S.N. Ahmed, P.K. Rao and V. Dakshinamurti, 1986. Plant products toxic to the citrus nematode *Tylenchulus semipenetrans* Cobb). Int. Nematol. Network Newslett., 3: 14-15.
 13. Rhode, R.A., 1960. Acetylcholinesterase in plant parasitic nematodes and an anti-cholinesterase from asparagus. Proc. Helminthol. Soc. Wash., 27: 121-127.
 14. Nelmes, A.J., 1970. Behavioral responses of *Heterodera rostochiensis* larvae to aldicarb and its sulfoxide and sulfone. J. Nematol., 2: 223-227.
 15. Kaliram, D. and C. Gupta, 1980. A note on efficacy of fresh neem leaf extract in the control of *Meloidogyne javanica* infesting chickpea *Cicer arietinum*. Ind. J. Nematol., 10: 96-98.
 16. Goswami, B.K. and K. Vijayalakshmi, 1986. Efficacy of some indigenous plant materials and non-edible oilseed cakes against *Meloidogyne incognita* on tomato. Indian J. Nematol., 16: 280-281.
 17. Nandal, S.N. and D.S. Bhatti, 1987. Effect of some shrub extracts on penetration and gall formation by *Meloidogyne javanica* in brinjal. Nematol. Medit., 15: 159-162.
 18. Kumari, R., K.K. Verma, K.S. Dhindsa and D.S. Bhatti, 1986. *Datura*, *Ipomea* and *Lawsonia* as control of *Tylenchulus semipenetrans* and *Anguina tritici*. Indian J. Nematol., 16: 236-240.
 19. Giebel, G., 1982. Mechanism of resistance to plant nematodes. Ann. Rev. Phytopathol., 20: 257-279.
 20. Kast, W.K., 1985. Wirkung alternativer spritzfolgen auf pilzliche Schaderreger bei Reben. Gesunde Pfl., 37: 494-501.
 21. Bunt, J.A., 1975. Effect and mode of action of some systemic nematicides. Meded. Landb-Hogesh. Wagenigen, 75: 1-128.
 22. Siddiqui, M.A. and M.M. Alam, 1987. Control of plant parasitic nematodes by intercropping with *Tagetes minuta*. Nematol. Medit., 15: 205-211.
 23. Siddiqui, M.A. and M.M. Alam, 1987. Efficacy of seed dressing with extracts of neem and persian lilak against *Meloidogyne incognita* and *Rotylenchulus reniformis*. Nematol. Mediterranea, 15: 399-403.
 24. Siddiqui, M.A. and M.M. Alam, 1988. Studies on the nemato-toxicity of root exudates of certain species of *Tagetes*. Indian J. Nematol., 18: 335-337.
 25. Hussain, S.I., R. Kumar, T. Khan and A. Titor, 1984. Effect of root dip treatment of egg plant seedlings with plant extracts, nematicides, oil cake extracts and anthelmintic drugs on plant growth and root knot development. Pak. J. Nematol., 2: 79-83.
 26. Bell, A.A., 1981. Biochemical mechanisms of disease resistance. Annu. Rev. Plant Physiol., 32: 21-81.



This document was created with the Win2PDF "print to PDF" printer available at <http://www.win2pdf.com>

This version of Win2PDF 10 is for evaluation and non-commercial use only.

This page will not be added after purchasing Win2PDF.

<http://www.win2pdf.com/purchase/>